Trade-off between fast growth and the ossification process in common carp (Cyprinus carpio)

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ABSTRACT

The relationship between growth rate and ossification is not consistent among fish. In some species, fast-growing individuals ossify earlier, while in other species, a trade-off between growth rate and ossification exists. This research was conducted to test the hypothesis about the existence of a trade-off mechanism in common carp offspring. For this purpose, specimens from two groups of common carp offspring (96 in total) with different growth rates were sampled at multiple time points between 5 and 29 days post-hatching. The standard length was measured and presented in correlation with the days post-hatching. To evaluate the skeletal system development, whole specimens were fixed in 10% neutral buffered formalin and stained for bone and cartilage. Particular emphasis was placed on the onset of ossification in the clearly visible bones of the head, vertebral column, and fins. According to our findings, larger, fast-growing specimens ossify earlier. The hypothesis on the existence of a trade-off between fast growth and the onset of ossification in common carp was not confirmed in our study.

Key words: trade-off; fast growth; ossification; common carp

Introduction

Growth rate, body weight and disease resistance are important criteria for selection programs in aquaculture (HULATA, 1995). Scientists are concerned that rapid growth could have different fund costs, e.g. reduced swimming performance (GREGORY and WOOD, 1988; LI et al. 2007;
The relationship between growth rate and ossification is not consistent in fish (KAVANAGH and ALFORD, 2003; BURDI, 2010; DESCHAMPS and SIRE, 2010). For instance, ARENDT and WILSON (2000) reported the potential cost of rapid growth in two populations of pumpkinseed (Lepomis gibbosus). The authors pointed out that the fish from the fast-growing population showed an average delay of 2 days in the onset of cranial bone ossification. Furthermore, they suggested that “such a delay in ossification may have fitness effects through feeding ability, swimming ability and (or) defense against predators”. However, opposite results were obtained in the study by KUŽIR et al. (2018) on the relationship of body size and osteological development in two common carp strains, Poljana mirror carp (PMC) and Poljana scaly carp (PSC). The authors stated that juveniles of PMC, at the same point of sampling (4, 15, 17, 19, 22, 24, and 26-days post-hatching (DPH)), achieved greater mass and length than juveniles of PSC. The osteological development had the same pattern in both groups, but with a delay in PSC. In conclusion, the authors deny any trade-off between fast growth and the ossification process in the common carp strains studied.

This study will present data about the onset of ossification in correlation to body size at certain ontogenetic points in cultured common carp. The main goal of this research is to test the hypothesis on the existence of a trade-off mechanism between the ossification process and growth at the intraspecific level. The long-term purpose of the current study is to give an insight into the ontogenetic development of common carp, which can be useful for carp aquaculture.

Materials and methods

The research was conducted at a fish farm in Croatia during a regular carp breeding program, as a part of non-experimental clinical veterinary practice. For the purpose of artificial propagation, two females, marked “S” and “C”, were selected and hand stripped to obtain eggs. The fertilization of eggs was performed in separate plastic bowls using mixed milt from two males. The fertilized eggs were incubated in separate incubation jars until they hatched. For the first five days, the larvae were kept in separate larvae containers and then stocked in two small nursery ponds (2 ha each) at a rate of 330,000/ha. Feeding of fry in the nursery ponds was based on natural food, while additional feed was introduced from 9th DPH (50% fish meal, 50% wheat flour, enriched with vitamin and mineral mixtures). Offspring from both groups (S and C) were sampled at multiple time points, starting from day 5 after hatching. The first sample was taken on the 5th DPH, on the day of stocking in the nursery ponds. Further sampling was conducted at 17, 22, 24, 26, and 29 DPH. Every sample contained 8 randomly selected specimens. After sampling and before taking measurements, the fish were euthanized by immersion in a buffered solution of MS-222 (Sigma-Aldrich, St. Louis, MO, USA). Standard length (SL) was measured and used for correlation with DPH. SL was measured from the most anterior part of the head to the posterior edge of hypural elements (ROJO, 1991). Statistical analysis was performed in STATISTICA v.13.5 (Statistica, Inc., 2018). For both groups, the mean and standard deviation (SD) was calculated. Differences between groups were analyzed with Mann - Whitney U test, with a significance level of P<0.05. To evaluate the skeletal system development, whole specimens were fixed in 10% neutral buffered formalin and stained for bone and cartilage, according to DINGERKUS and UHLER (1977). This double staining method allowed simultaneous detection of cartilaginous elements (alcian blue) as well as calcified structures (alizarin red). Stained specimens were visualized and photographed under an Olympus SZX7 stereoscopic microscope. Clearly visible bony elements and the onset of ossification process was described for the frontal, parietal, supraoccipital,
parasphenoid, basisphenoid, basioccipital, premaxillary, maxillary, dentary, articular, quadrate, hyomandibular and ceratohyal bones, the opercle, cleithrum, abdominal and caudal vertebrae, and the dorsal, caudal, anal, pelvic and pectoral fins. The study was approved by the Institutional Ethics Committee (Faculty of Veterinary Medicine, University of Zagreb).

Results

A total of 96 specimens were examined. The mean and standard deviation of body length for both groups are shown in Table 1, while the correlation of the standard length with days post-hatching is presented in Fig. 1.

In group S the larvae were $5.80 \pm 0.31$ mm SL at 5 DPH and there was no sign of the onset of ossification (Fig. 2).

The ossification process is visible in some head bones (parasphenoid, basisphenoid, ceratohyal, cleithrum) at 17 DPH (SL $10.49** \pm 0.48$ mm). At that stage, the very beginning of ossification is visible on the outer edges of the first 8 abdominal vertebrae (Fig. 3a). At 22 DPH (SL $16.24** \pm 1.67$ mm) ossification started in the basioccipital, premaxillary, maxillary, dentary, articular, quadrate, and hyomandibular bones as well as the opercle. At this stage, ossification was also noticed in the caudal vertebrae, especially in the last four, where the bodies were fully ossified. At the same stage, the onset of ossification is visible in the hypural bones of the caudal fin. At 24 DPH (SL $17.98** \pm 2.23$ mm), ossification is also visible in the dorsal fin, while at 26 DPH (SL $21.11** \pm 4.15$ mm) ossification had started in the anal, pelvic and pectoral fins. At 29 DPH (SL $28.98* \pm 2.27$ mm), and three bones on the head, the frontal, parietal and supraoccipital, had started to ossify. The process of ossification was either evolving or completed on the other investigated elements, except for the fins where ossification was still not complete.

![Fig. 1. Standard length (mm) of common carp offspring at the sampling time points (DPH)](image)

![Fig. 2. Five DPH old larva from group S. The existing skeleton is composed of cartilage. Scale bar = 1 mm.](image)

Table 1. Total number, mean and standard deviation of body length (SL) in mm for S and C group

<table>
<thead>
<tr>
<th>Group</th>
<th>N</th>
<th>5 DPH (Mean ± SD)</th>
<th>17 DPH (Mean ± SD)</th>
<th>22 DPH (Mean ± SD)</th>
<th>24 DPH (Mean ± SD)</th>
<th>26 DPH (Mean ± SD)</th>
<th>29 DPH (Mean ± SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>S</td>
<td>8</td>
<td>5.80 ± 0.31</td>
<td>10.49** ± 0.48</td>
<td>16.24** ± 1.67</td>
<td>17.98** ± 2.23</td>
<td>21.11** ± 4.15</td>
<td>28.98* ± 2.27</td>
</tr>
<tr>
<td>C</td>
<td>8</td>
<td>5.87 ± 0.28</td>
<td>13.39** ± 0.44</td>
<td>24.67** ± 1.02</td>
<td>28.06** ± 2.00</td>
<td>28.81** ± 2.10</td>
<td>31.85* ± 1.55</td>
</tr>
</tbody>
</table>

* Values in the same column are significantly different, *(P<0.05), **(P<0.01).
The ossification process is visible in some head bones (parasphenoid, basisphenoid, ceratohyal and cleithrum) at 17 DPH (SL 10.49** ± 0.48 mm). At that stage, the very beginning of ossification is visible on the outer edges of the first 8 abdominal vertebrae.

In group C at 5 DPH the larvae were 5.87 ± 0.28 mm SL and there was no sign of ossification. The ossification process is visible in most of the head bones (parasphenoid, basisphenoid, basioccipital, premaxillary, maxillary, dentary, hyomandibular, ceratohyal, opercle, and cleithrum) at 17 DPH (SL 13.39** ± 0.44 mm) (Fig. 3b). At this stage, ossification is also visible in the abdominal and caudal vertebrae. At 22 DPH (SL 24.67** ± 1.02 mm), ossification had started in the supraoccipital, articular and quadrate bones. The first sign of ossification was noticed in the dorsal and caudal fins (Fig. 4a and 4b). At 24 DPH (SL 28.06** ± 2.00 mm), a new sign of ossification is visible in the head, the frontal and parietal bones, as well as in the anal, pelvic and pectoral fins. Ossification was in progress or completed by 26 DPH, (SL was 28.81** ± 2.10 mm) in all investigated elements of the head and vertebral column. At 29 DPH, (SL 31.85* ± 1.55 mm) ossification had finished in the anal fin while the other fins still had cartilaginous parts.

The onset of ossification per day post-hatching for both groups is shown in Fig. 5.
<table>
<thead>
<tr>
<th>Bones</th>
<th>DPH</th>
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<tbody>
<tr>
<td></td>
<td>5</td>
</tr>
<tr>
<td>Frontal</td>
<td></td>
</tr>
<tr>
<td>Parietal</td>
<td></td>
</tr>
<tr>
<td>Supraoccipital</td>
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<tr>
<td>Parasphenoid</td>
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<td>Basisphenoid</td>
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<td>Basioccipital</td>
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<td>Premaxillary</td>
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<tr>
<td>Maxillary</td>
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<tr>
<td>Dentary</td>
<td></td>
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<tr>
<td>Articular</td>
<td></td>
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<tr>
<td>Quadrato</td>
<td></td>
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<tr>
<td>Hyomandibular</td>
<td></td>
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<tr>
<td>Ceratohyal</td>
<td></td>
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<tr>
<td>Opercle</td>
<td></td>
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<tr>
<td>Cleithrum</td>
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<tr>
<td>Abdominal vertebrae</td>
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<tr>
<td>Caudal vertebrae</td>
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<tr>
<td>Dorsal fin</td>
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<tr>
<td>Caudal fin</td>
<td></td>
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<tr>
<td>Anal fin</td>
<td></td>
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<td>Pelvic fin</td>
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<tr>
<td>Pectoral fin</td>
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</table>

Fig. 5. The onset of the ossification process of clearly visible elements in group S (gray bars) and group C (black bars)
Discussion

Despite the fact that the common carp is one of the oldest cultured and most domesticated fish in the world (WOHLFARTH, 1995; BAKOS and GORDA, 2001), its ontogenetic processes have not been fully elucidated, and there is a great variability and fragmentation in the data published. A possible reason for this could be the involvement of numerous insufficiently researched external (non-genetic) and internal (genetic, micro environmental) factors that modulate morphogenesis (BOGLIONE et al., 2013). According to OSSE and VAN DEN BOOGAART (1995) and MABEE et al. (2002) development of morphological structures is strongly correlated with functional requirements. This conclusion is in accordance with the results of our study.

At 5 DPH, there was no statistical difference in the values of SL between groups S and C. In both groups, the existing skeleton was made of cartilage and there was no sign of ossification. The same was previously described by ITIZAWA (1963) and PASHINE and MARATHE (1977). From 17 DPH to 26 DPH, the values of SL differed statistically between the groups (Table 1) and, importantly, a variation in the onset of ossification is present (Fig. 2). In both groups, ossification begins at 17 DPH in the parasphenoid, basisphenoid, ceratohyal and cleithrum, which could be explained by the functional requirements at this life stage (MOSHAYEDI et al., 2016). The parasphenoid and basisphenoid form the base of the cranium, give firmness to the skull, and act as an attachment site for some masticatory muscles. The ceratohyal has the main role in articulation with branchiostegal rays. The cleithrum extends from the pectoral fin and forms the posterior edge of the gill chamber. At 17 DPH, in group C, ossification was also present in the basisioccipital, premaxillary, maxillary and dentary, as well as the hyomandibular and opercle. PASHINE and MARATHE (1979) described a similar onset of ossification, but in much smaller common carp larvae (total length 7.5 mm). At 22 DPH, ossification was noticed in both groups in the articular and quadrate bones. In S group, it was visible in the basisioccipital, premaxillary, maxillary, dentary, hyomandibular and opercle. Additionally, in C group ossification started in the supraoccipital.

Although the ossification sequence in the head is similar, a delay was visible in group S. At 24 DPH the frontal and parietal started to ossify in C group, while in S group these two bones, as well as the supraoccipital, only showed signs of ossification at 29 DPH.

In the vertebral column, the first sign of ossification was noticed in the abdominal vertebrae at 17 DPH in both groups. Ossification started on the outer parts of the first three vertebral bodies, and proceeded backwards. The caudal vertebrae started to ossify at 17 DPH in group C, but at 22 DPH in group S. The last three caudal vertebrae and urostyle ossified first, and the ossification proceeds forward. Considering the time and onset of ossification of the vertebrae, the results of the present study do not coincide with those of ITIZAWA (1963). ITIZAWA (1963) reported that the ossification sequences of the vertebral column occur much earlier in common carp larvae (5 DPH/SL 7.0 mm), and with a dissimilar pattern. The author placed special emphasis on the first three vertebral bodies that ossify only after the backwards proceeding ossification has reached the 15th vertebra. Furthermore, ITIZAWA (1963) pointed out that the ossification sequence in the caudal region of the vertebral column is the most complicated. According to him, the whole process of vertebral column ossification is completed at 15 DPH (SL 10.9 mm). SŁOMIŃSKA and JEZIERSKA (2000) recorded the end of ossification in the vertebral column of common carp at around 40 DPH, which is 25 days later than ITIZAWA (1963) stated and 11 days later than was observed in this study.

Ossification of the fins started at 22 DPH. In both groups, the dorsal fin ossified first. Simultaneously, in group C ossification was also noticed in the caudal fin. In group S the ossification process started in the caudal fin at 24 DPH. Ossification of the anal and pectoral fins in group C was noticed at 24 DPH, while in S group it occurred at 26 DPH. In both groups, the pelvic fins started to ossify at 26 DPH. The same order was described by ITIZAWA (1963), but the starting point was much earlier (12 DPH/SL 9.3 mm).
Conclusion

The onset of ossification is similar in both studied common carp groups, but not completely identical. Inconsistency with data from the literature was also found. Furthermore, we confirmed that in common carp the sequence of ossification is not determined by age (DPH). Significantly, larger, fast-growing specimens ossify earlier. No trade-off between fast growth and onset of ossification was confirmed in our study.

References


BAKOS, J., S. GORDA (2001): Genetic resources of common carp at the Fish Culture Research Institute, Szarvas, Hungary. FAO, Rome.


S. Kužir et al.: Growth and ossification in common carp

DOI: 10.1242/jeb.148056

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SAŽETAK
Odnos između brzine rasta i procesa okoštavanja u riba nije ujednačen. U nekih vrsta te su vrijednosti u pozitivnoj korelaciji, dok je u drugih vrsta prisutan svojevrsni kompromis (engl. trade-off), brži rast negativno utječe na proces okoštavanja. Cilj ovog istraživanja je provjeriti hipotezu o postojanju negativne korelacije između brzine rasta i procesa okoštavanja u šarana (Cyprinus carpio). U tu svrhu, jedinke šarana podrijetlom iz dvije skupine s različitom brzinom rasta (ukupno 96 jedinki) uzorkovane su između 5. i 29. dana nakon valjenja. Mjerena je standardna dužina i prikazana u korelaciji s brojem dana poslije valjenja. U svrhu procjene razvoja koštanog sustava, uzorci su fiksirani u 10% neutralnom formalinu i obojeni metodom za prikaz kosti i hrskavice. Poseban naglasak stavljen je na proces okoštavanja u jasno vidljivim kostima glave, kralježnice i peraja. Prema rezultatima našeg istraživanja, u brzorastućih jedinki proces okoštavanja je nastupio ranije. U ovom radu opovrgnuta je hipoteza o postojanju negativne korelacije između brzine rasta i procesa okoštavanja.

Ključne riječi: trade-off; brzina rasta; okoštavanje; šaran