Detection of *Chlamydia psittaci* genotypes in fecal samples of homing pigeons in Croatia

Danijela Horvatek Tomić^{1*}, Karine Laroucau², and Estella Prukner-Radovčić¹

¹Department of Poultry Diseases with Clinic, Faculty of Veterinary Medicine University of Zagreb, Zagreb, Croatia

²Bacterial Zoonoses Unit, French Agency for Food, Environmental and Occupational Health Safety (Anses), Maisons-Alfort, France

HORVATEK TOMIĆ, D., K. LAROUCAU, E. PRUKNER-RADOVČIĆ: Detection of *Chlamydia psittaci* genotypes in fecal samples of homing pigeons in Croatia. Vet. arhiv 83, 201-209, 2013.

ABSTRACT

In this investigation fecal samples from a total of 232 homing pigeon lofts, belonging to both racing and breeding pigeons, were examined by specific real-time PCR to reveal the presence of Chlamydiaceae, and randomly collected Chlamydia positive samples were genotyped by two different molecular methods; C. psittaci-specific restriction fragment length polymorphism (RFLP) and Multiple-Locus Variable number tandem repeat Analysis (MLVA), in order to obtain relevant information about the prevalence of different genotypes, and to reveal the potential threat to humans that come into close contact with homing pigeons. Chlamydiosis in birds manifests as an acute, unapparent, sub-clinical, and chronic disease, but frequently also as an asymptomatic infection, which represents an additional danger for public health. Currently, nine different genotypes of C. psittaci have been generally accepted, based on PCR amplification, as well as an additional six provisional genotypes, based on DNA microarray. The predominant serotype/genotype in pigeons is B, but also other genotypes were discovered, such as A, C and D, as well as mixed infection. Out of 232 examined samples, 30 (12.9%) of them were PCR positive. DNA from six random positive fecal samples was further examined and the presence of C. psittaci was confirmed in all of them. According to RFLP genotyping, from 6 C. psittaci positive samples, 4 of them belonged to genotype B, and 2 strains remained untyped, due to the low concentration of DNA isolated. Regarding the MLVA typing, the pattern assigned as 22146334 could be described as a classical "pigeon" MLVA pattern. The presence of 12.9% homing pigeons positive to Chlamydiaceae by PCR clearly demonstrates the presence of zoonotic microorganisms and a possible risk for pigeon fanciers that come in close contact with the birds.

Key words: genotyping, Chlamydia psittaci, pigeons

ISSN 0372-5480
Printed in Croatia

^{*}Corresponding author:

Dr. Danijela Horvatek Tomić, DVM, Department of Poultry Diseases with Clinic, Faculty of Veterinary Medicine University of Zagreb, Heinzelova 55, 10000 Zagreb, Croatia, Phone: +385 1 2390 281; Fax: +385 1 2390 280; E-mail: horvatek@yef.hr

Introduction

Chlamydia (formerly Chlamydophila) psittaci (C. psittaci) infects a wide range of birds all around the world and has been detected in altogether 465 avian species (KALETA and TADAY, 2003). Avian chlamydiosis manifests as an acute, unapparent, sub-clinical, and chronic disease, but frequently also as an asymptomatic infection (ANDERSEN and VANROMPAY, 2003; SUDLER et al., 2004). On the basis of the immunofluorescent reaction of the major outer membrane protein (MOMP) with a panel of specific monoclonal antibodies, C. psittaci have been classified into six serovars: A, B, C, D, E and F (ANDERSEN, 1991a, b; VANROMPAY et al., 1993). Nowadays, based on PCR amplification, each serovar could be assigned to an equivalent genotype, so nine different genotypes have been generally accepted: the six already mentioned that predominantly occur in birds, M56 in rodents, WC in cattle (VANROMPAY et al., 1997), and the most recently described, E/B (GEENS et al., 2005). According to SACHSE et al. (2008), adjustments to the present genotypes scheme should be made by introduction of six provisional genotypes, as detected by DNA microarray.

Urban pigeons are often infected with *C. psittaci* all over the world, and a high percentage of seropositive, but also infected birds can be found, respectively (GAFFURI et al., 2000; DOVČ et al., 2004; MAGNINO et al., 2009). The predominant serotype/genotype in pigeons is B (HEDDEMA et al., 2006a; GASPARINI et al., 2011; GEIGENFEIND et al., 2012), but also other genotypes have been discovered, as A (VANROMPAY et al., 1997; SACHSE et al., 2012), C and D (DICKX et al., 2010), E/B (GEENS et al., 2005), mixed infection (GEIGENFEIND et al., 2012), or provisional genotypes (SACHSE et al., 2012).

In the Republic of Croatia, according to state laws, all avian flocks and birds in breeding and transport must be tested for avian chlamydiosis. The investigations implemented in Croatia so far have revealed the frequent presence of *Chlamydia* in urban pigeons (PAVLAK et al., 2000; PRUKNER-RADOVČIĆ et al., 2005). Racing and breeding pigeons, also known as homing pigeons, are both very popular in Croatia. Lofts are mostly kept outdoors, and as such, contact with infected urban pigeons or other birds is possible. Samples collected from different bird species (mostly fecal samples, swabs or organs) are examined by using several diagnostic methods, however, particular classification and identification is completed by conventional PCR and real-time PCR (HEWINSON et al., 1997; EHRICHT et al., 2006; PANTCHEV et al., 2008). Determining of the *C. psittaci* genotypes is especially important from the public health point of view, since it is well known that certain genotypes can cause severe infection in humans (especially A and D), and other (mainly B) just mild respiratory symptoms (DICKX et al., 2010) or no clinical signs at all (SACHSE et al., 2012). Numerous cases of zoonoses caused by *C. psittaci*, linked to pigeons, were described by HAAG-WACKERNAGEL and MOCH (2004).

Genotyping techniques have already been used for the typing of avian *C. psittaci* strains by SAYADA et al. (1995). Recently, a novel approach, based on the finding of tandem repeats in DNA (Multiple-Locus Variable number tandem repeat, MLVA) has been applied for genotyping of *C. psittaci*. This method targets eight distinct genomic areas, dispersed throughout the genome (LAROUCAU et al., 2008). More recently, DNA microarray technology was also used for typing of *C. psittaci* and was shown to discriminate among already known genotypes and identify so far untyped strains (SACHSE et al., 2008; SACHSE et al., 2009).

In this investigation, fecal samples belonging to both racing and breeding pigeon flocks were examined by *Chlamydiaceae*-specific real-time PCR (targeting the 23S rRNA gene) and *C. psittaci*-specific real-time PCR (targeting the *ompA* gene), and randomly collected *Chlamydia* positive samples were genotyped by two different molecular methods; *C. psittaci*-specific RFLP and MLVA scheme, in order to obtain relevant information about the prevalence of all, up to now, known genotypes, and to reveal the potential threat to humans that are in close contact with homing pigeons.

Materials and methods

Fecal samples from a total of 232 homing pigeon lofts, originating from both racing and breeding pigeons of different ages and sizes of population, were taken by the official veterinary authorities in all 21 counties of the Republic of Croatia and in the City of Zagreb for routine diagnosis of *Chlamydiaceae*, according to state law, and send to the Laboratory for Chlamydia at the Department of Poultry Disease and Clinic, of the Faculty of Veterinary Medicine, University of Zagreb. One pooled fecal sample, belonging to approximately 10% of the pigeon population was taken for each loft. The DNA was extracted using a GenElute Mammalian Genomic DNA Miniprep Kit (Sigma-Aldrich, USA), according to the manufacturer's instructions, and examined by *Chlamydiaceae*-specific real-time PCR, as described by EHRICHT et al. (2006).

Further, randomly chosen DNA samples, belonging to 6 different pigeons lofts, were examined at Unité Zoonoses Bactériennes, Anses Maisons-Alfort, France, by *C. psittaci*-specific real-time PCR, as described by PANTCHEV et al. (2008), and genotyped by alternate molecular tools - MLVA and RFLP. Genotyping by RFLP, based on restriction enzymes *Alu*I and *Mbo*II, was performed as described by SAYADA et al. (1995). Fragments were separated on a 2% NuSieve agarose gel and visualized under a UV light. MLVA were conducted as described by LAROUCAU et al. (2008), by using 8 sets of primers: ChlaPsi_280, ChlaPsi_480, ChlaPsi_605, ChlaPsi_810, ChlaPsi_222, ChlaPsi_281, ChlaPsi_929 and ChlaPsi_1788.

Results

Table 1. Genotyping of pigeon's C. psittaci strains (n 6).

| | | | C. psittaci genotyping | |
|-----------|----------------------------|---------------------------|------------------------|----------|
| Sample ID | 23S-rtPCR Chlamydiaceae | ompA rtPCR C. psittaci | PCR-RFLP pattern | MLVA |
| 1 | + | + | genotype B | 22146334 |
| 2 | + | + | genotype B | 22146334 |
| 3 | + | + | nec* | nec |
| 4 | + | + | genotype B | 22146334 |
| 5 | + | + | genotype B | 22146334 |
| 6 | + | + | nec | nec |

^{*}nec - not enough concentrated DNA

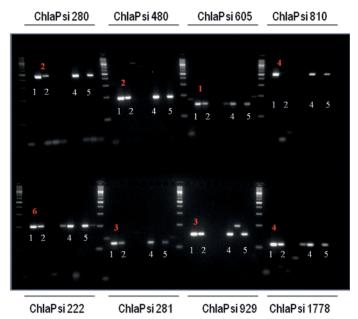


Fig. 1. MLVA of 4 pigeon's C. *psittaci* strains. Numbers in white corresponds with the sample ID number (1, 2, 4 and 5) and numbers in red with the MLVA pattern (22146334). A 100 bp ladder is run on each side of a group of samples. Samples numbered as 3 and 6 were not included since there was not enough DNA. Primers used are signed as ChlaPsi 280, 480, 605, 810, 222, 281, 929 and 1778.

Altogether 232 fecal samples, from racing and breeding pigeon lofts were examined by real-time PCR, to reveal the presence of *Chlamydiaceae*, according to Croatian laws (ANONYM., 2011). Of these, 30 (12.9%) fecal samples were shown to be PCR positive. DNA from six random positive fecal samples were further examined and the presence of C. *psittaci* was confirmed in all of them (Table 1).

According to RFLP genotyping, from six *C. psittaci* positive samples, four of them belonged to genotype B, and two strains remained untyped, due to the low concentration of extracted DNA. Regarding the MLVA typing, the pattern assigned as 22146334 (Fig. 1), described as a classical "pigeon" MLVA pattern, was found in all four examined samples.

Discussion

The detection rate of *Chlamydia* found in the fecal samples of homing pigeons in Croatia is in concordance with the findings of other authors (DICKX et al., 2010; SACHSE et al., 2012). Recently it was also described that the detection rate of *Chlamydia* in pigeon fecal samples by PCR could be even better than in swabs, due to the combining of feces from possibly high shedders in one representative sample belonging to an entire loft (SACHSE et al., 2012). This could confirm the results of this study, as from only two samples was the DNA yield too low and genotyping was not possible.

Genotyping, performed by two different methods, revealed the presence of genotype B in all examined samples. The results of RFLP typing were consistent with MLVA schemes, but with one difference - MLVA did not provide the distinction between B and E genotypes. This apparent discrepancy probably reflects the fact that none of the markers chosen in this study target *omp*A (LAROUCAU et al., 2008), as were in RFLP. The results of genotyping are in conclusion with the majority of other authors, who also reported that pigeons predominantly carry genotype B (VANROMPAY et al., 1997; HEDDEMA et al., 2006b; PANNEKOEK et al., 2010; GASPARINI et al., 2011; GEIGENFEIND et al., 2012), but other genotypes were also discovered (VANROMPAY et al., 1997; DICKX et al., 2010; GEIGENFEIND et al., 2012). The genotype B, originating from nonpsittacine birds, has always been described as having low or medium virulence, but there are also some reports that it may be the cause of human disease (HEDDEMA et al., 2006b)

Homing and urban pigeons often shed chlamydiae intermittently in the absence of clinical signs, which makes diagnosis very cumbersome. On the other hand, shedding of this pathogen through feces, and respiratory and conjunctival secretions, increases the risk of transmission of *C. psittaci* to other animals and to humans (ANDERSEN and VANROMPAY, 2003; HARKINEZHAD et al., 2009; VAN DROOGENBROECK et al., 2009). According to HARKINEZHAD et al. (2009) up to 7% of apparently healthy people, coming in close contact with pigeons, were positive by PCR to *C. psittaci* or had antibodies

against it. In the study conducted by DICKX et al. (2010) the number of positive persons was even higher - 12.5%.

Previous investigations conducted in the City of Zagreb (the Capital of Croatia) revealed approximately 15% of urban pigeons positive to *C. psittaci* (PRUKNER-RADOVČIĆ et al., 2005; HORVATEK et al., 2007). The current investigation revealed the presence of 12.9% homing pigeons positive to *Chlamydiaceae* by PCR, which clearly demonstrates the presence of zoonotic microorganisms and a possible risk for pigeon fanciers that come into close contact with the birds. Homing pigeons positive to *C. psittaci* are treated with antibiotics, but due to the possibility of development of resistant strains, as described by DUGAN et al. (2004), it is obligatory to treat the infected animals under strict veterinary supervision.

In this investigation the identity of the species of all Chlamydiaceae-positive samples was not determined, and according to other authors not only *C. psittaci*, but also *C. abortus*, *C. pecorum*, or *C. trachomatis* can be found in pigeons and other birds (PANTCHEV et al., 2009; SACHSE et al., 2012), so further investigations will be needed to clearly identify positive samples.

Acknowledgements

The investigation was partly supported by the Ministry of Science and Technology of the Republic of Croatia, Grants No. 053-0531863-1857

References

- ANDERSEN, A. A. (1991a): Comparison of avian *Chlamydia psittaci* isolates by restriction endonuclease analysis and serovar-specific monoclonal antibodies. J. Clin. Microbiol. 29, 244-249
- ANDERSEN, A. A. (1991b): Serotyping of *Chlamydia psittaci* isolates using serovar-specific monoclonal antibodies with the microimmunofluorescence test. J. Clin. Microbiol. 29, 707-711.
- ANDERSEN, A. A., D. VANROMPAY (2003): Avian chlamydiosis (psittacosis, ornithosis). In: Diseases of Poultry 11th ed. (Saif, Y. M., Ed.), Iowa State University Press, Iowa, pp. 863-879.
- ANONYMOUS (2011): Naredba o mjerama zaštite životinja od zaraznih i nametničkih bolesti i njihovom financiranju u 2011. godini. NN 1/11. (Order on measures to protect animals from infections and parasitic diseases and financing thereof in 2011. Official Gazette no. 1/11).
- DICKX, V., D. S. BEECKMAN, L. DOSSCHE, P. TAVERNIER, D. VANROMPAY (2010): *Chlamydophila psittaci* in homing and feral pigeons and zoonotic transmission. J. Med. Microbiol. 59, 1348-1353.

- DOVČ, A., O. ZORMAN-ROJS, A. VERGLES RATAJ, V. BOLE-HRIBOVŠEK, U. KRAPEŽ, M. DOBEIC (2004): Health status of free-living pigeons (*Columba livia domestica*) in the city of Ljubljana. Acta Vet. Hung. 52, 219-226.
- DUGAN, J., D. D. ROCKEY, L. JONES, A. A. ANDERSEN (2004): Tetracycline resistance in Chlamydia suis mediated by genomic islands inserted into the chlamydial inv-like gene. Antimicrob. Agents Chemother. 48, 3989-3995.
- EHRICHT, R., P. SLICKERS, S. GOELLNER, H. HOTZEL, K. SACHSE (2006): Optimized DNA microarray assay allows detection and genotyping of single PCR-amplifiable target copies. Mol. Cell. Probes 20, 60-63.
- GAFFURI, A., C. GARBARINO, T. CONSOLI, M. CARRARA, D. BROUSSARD, V. PIETRA, S. MAGNINO, F. PATERLINI (2000): Study on the health status of pigeons in Bergamo province. Sel. Vet. 8-9, 827-830.
- GASPARINI, J., N. ERIN, C. BERTIN, L. JACQUIN, F. VORIMORE, A. FRANTZ, P. LENOUVEL, K. LAROUCAU (2011): Impact of urban environment and host phenotype on the epidemiology of *Chlamydiaceae* in feral pigeons (*Columba livia*). Environ. Microbiol. 13, 3186-3193.
- GEENS, T., A. DESPLANQUES, M. VAN LOOCK, B. M. BONNER, E. F. KALETA, S. MAGNINO, A. A. ANDERSEN, K. D. EVERETT, D. VANROMPAY (2005): Sequencing of the *Chlamydophila psittaci* ompA gene reveals a new genotype, E/B, and the need for a rapid discriminatory genotyping method. J. Clin. Microbiol. 43, 2456-2461.
- GEIGENFEIND, I., D. VANROMPAY, D. HAAG-WACKERNAGEL (2012): Prevalence of *Chlamydia psittaci* in the feral pigeon population of Basel, Switzerland. J. Med Microbiol. 61, 261-265.
- HAAG-WACKERNAGEL, D., H. MOCH (2004): Health hazards posed by feral pigeons. J. Infect. 48, 307-313.
- HARKINEZHAD, T., K. VERMINNEN, M. DE BUYZERE, E. RIETZSCHEL, S. BEKAERT, D. VANROMPAY (2009): Prevalence of *Chlamydophila psittaci* infections in a human population in contact with domestic and companion birds. J. Med. Microbiol. 58, 1207-1212.
- HEDDEMA, E. R., S. TER SLUIS, J. A. BUYS, C. M. J. E. VANDENBROUCKE-GRAULS, J. H. VAN WIJNEN, C. E. VISSER (2006a): Prevalence of *Chlamydophila psittaci* in fecal droppings from feral pigeons in Amsterdam, the Netherlands. Appl. Environ. Microbiol. 72, 4423-4425.
- HEDDEMA, E. R., E. J. VAN HANNEN, B. DUIM, C. M. J. E. VANDENBROUCKE-GRAULS, Y. PANNEKOEK (2006b): Genotyping of *Chlamydophila psittaci* in human samples. Emerg. Inf. Dis. 12, 1989-1990.
- HEWINSON, R. G., P. C. GRIFFITHS, B. J. BEVAN, S. E. S. KIRWAN, M. E. FIELD, M. J. WOODWARD, M. DAWSON (1997): Detection of *Chlamydia psittaci* DNA in avian clinical samples by polymerase chain reaction. Vet. Microbiol. 54, 155-166.
- HORVATEK, D., Ž. GOTTSTEIN, I. CIGLAR GROZDANIĆ, H. MAZIJA, K. VLAHOVIĆ, E. PRUKNER-RADOVČIĆ (2007): Urban pigeons in the City of Zagreb a possible source of

- *Chlamydophila psittaci* infection in human. Proceedings of the 5th Annual Workshop of COST Action 855 Animal Chlamydioses and Zoonotic Implications, 09-11 September, Pulawy, Poland, p. 33.
- KALETA, E. F., E. M. TADAY (2003): Avian host range of *Chlamydophila* spp. based on isolation, antigen detection and serology. Avian Pathol. 32, 435-461.
- LAROUCAU, K., S. THIERRY, F. VORIMORE, K. BLANCO, E. KALETA, R.HOOP, S. MAGNINO, D. VANROMPAY, K. SACHSE, G. S. A. MYERS, P. M. BAVOIL, G. VERGNAUD, C. POURCEL (2008): High resolution typing of *Chlamydophila psittaci* by multilocus VNTR analysis (MLVA). Infect. Genet. Evol. 8, 171-181.
- MAGNINO, S., D. HAAG-WACKERNAGEL, I. GREIGENFEIND, S. HELMECKE, A. DOVČ, E. PRUKNER-RADOVČIĆ, E. REŠIDBEGOVIĆ, V. ILESKI, K. LAROUCAU, M. DONATI, S. MARTINOV, E. F. KALETA (2009): Chlamydial infections in feral pigeons in Europe: Review of data and focus on public health implications. Vet. Microbiol. 135, 54-67.
- PANNEKOEK, Y., V. DICKX, D. S. BEECKMAN, K. A. JOLLEY, W. C. KEIJZERS, E. VRETOU, M. C. MAIDEN, D. VANROMPAY, A. VAN DER ENDE (2010): Multilocus sequence typing of *Chlamydia* reveals an association between *Chlamydia psittaci* genotypes and host species. Public Library of Science (PLoS One) 5, e14179, doi:10.1371/journal.pone.0014179
- PANTCHEV, A., R. STING, R. BAUERFEIND, J. TYCZKA, K. SACHSE (2008): New real-time PCR tests for species-specific detection of *Chlamydophila psittaci* and *Chlamydophila abortus* from tissue samples. Vet. J. 181, 145-150.
- PANTCHEV, A., R. STING, R. BAUERFEIND, J. TYCZKA, K. SACHSE (2009): New real-time PCR tests for species-specific detection of *Chlamydophila psittaci* and *Chlamydophila abortus* from tissue samples. Vet. J. 181, 145-150.
- PAVLAK, M., K. VLAHOVIĆ, J. GREGURIĆ, Ž. ŽUPANČIĆ, J. JERČIĆ, J. BOŽIKOV (2000): An epidemiologic study of *Chlamydia* sp. in feral pigeons (*Columba livia domestica*). Z. Jadgdwiss. 46, 84-95.
- PRUKNER-RADOVČIĆ, E., D. HORVATEK, Ž. GOTTSTEIN, I. CIGLAR GROZDANIĆ, H. MAZIJA (2005b): Epidemiological investigation on *Chlamydophila psittaci* in pigeons and free living birds in Croatia. Vet. Res. Commun. 29 (Suppl. 1), 17-21.
- SACHSE, K., K. LAROUCAU, H. HOTZEL, E. SCHUBERT, R. EHRICHT, P. SLICKERS (2008): Genotyping of *Chlamydophila psittaci* using a new DNA microarray assay based on sequence analysis of *omp*A genes. BMC Microbiol. 8, 63-75.
- SACHSE, K., K. LAROUCAU, F. VORIMORE, S. MAGNINO, J. FEIGE, W. MÜLLER, S. KUBE, H. HOTZEL, E. SCHUBERT, P. SLICKERS, R. EHRICHT (2009): DNA microarray-based genotyping of *Chlamydophila psittaci* strains from culture and clinical samples. Vet. Microbiol. 135, 22-30.
- SACHSE, K., S. KUEHLWIND, A. RUETTGER, E. SCHUBERT, G. ROHDE (2012): More than classical *Chlamydia psittaci* in urban pigeons. Vet. Microbiol. 157, 476-480.

- SAYADA, C., A. A. ANDERSEN, C. STOREY, A. MILON, F. EB, N. HASHIMOTO, K. HIRAI, J. ELION, E. DENAMUR (1995): Usefulness of *omp1* restriction mapping for avian *Chlamydia psittaci* isolate differentiation. Res. Microbiol. 146, 155-165.
- SUDLER, C., L. E. HOELZLE, I. SCHILLER, R. K. HOOP (2004): Molecular characterisation of chlamydial isolates from birds. Vet. Microbiol. 98, 235-241.
- VAN DROOGENBROECK, C., D. S. A. BEECKMAN, K. VERMINNEN, M. MARIEN, H. NAUWYNCK, L. DE THIBAULT DE BOESINGHE, D. VANROMPAY (2009): Simultaneous zoonotic transmission of *Chlamydophila psittaci* genotypes D, F and E/B to a veterinary scientist. Vet. Microbiol. 135, 78-81.
- VANROMPAY, D., A. A. ANDERSEN, R. DUCATELLE, F. HAESEBROUCK (1993): Serotyping of European isolates of *Chlamydia psittaci* from poultry and other birds. J. Clin. Microbiol. 31, 134-137.
- VANROMPAY, D., P. BUTAYE, C. SAYADA, R. DUCATELLE, F. HAESEBROUCK (1997): Characterization of avian *Chlamydia psittaci* strains using omp1 restriction mapping and serovar-specific monoclonal antibodies. Res. Microbiol. 148, 327-333.

Received: 19 March 2012 Accepted: 27 June 2012

HORVATEK TOMIĆ, D., K. LAROUCAU, E. PRUKNER-RADOVČIĆ: Genotipovi bakterije *Chlamydia psittaci* u golubova u Hrvatskoj. Vet. arhiv 83, 201-209, 2013.

Ukupno su bila pretražena 232 uzorka izmeta golubova na prisutnost bakterija porodice Chlamydiaceae pomoću za Chlamydiaceae specifične lančane reakcije polimerazom u stvarnom vremenu. Nasumično odabrani pozitivni uzorci pretraženi su i na prisutnost bakterije *Chlamydia psittaci* (*C. psittaci*), te su u slučaju pozitivnog nalaza i genotipizirani pomoću dvije različite molekularne metode - MLVA i RFLP, kako bi dobili svrsishodne informacije o učestalosti različitih genotipova, te otkrili potencijalnu prijetnju za zdravlje ljudi koji dolaze u bliži kontakt s golubovima. Klamidioza u ptica očituje se kao akutna, inaparentna, supklinička ili kronična bolest, ali često i kao asimptomatska zaraza, što predstavlja dodatnu opasnost za javno zdravstvo. Novijim istraživanjima, temeljenim na lančanoj reakciji polimerazom (PCR) nađeno je devet različitih genotipova C. psittaci. Prevladavajući serotip/genotip u golubova je B, ali su u njih nađeni i drugi genotipovi, kao što su A, C i D, te mješovite infekcije. Od 232 pregledana uzorka, njih 30 (12,9%) bilo je pozitivno na prisutnost bakterija iz porodice Chlamydiaceae. Nasumično izabrana DNK šest pozitivnih uzoraka dodatno je pretražena i na prisutnost C. psittaci, te je nalaz bio pozitivan u svih šest uzoraka. Pomoću PCR-RFLP metode, od 6 uzoraka pozitivnih na bakteriju C. psittaci, četiri su pripadala genotipu B, dok se dva uzorka nisu mogla pretražiti zbog niske koncentracije izdvojene DNK. Pomoću MLVA metode, četiri pretražena uzorka označena su oznakom 22146334, što se opisuje kao "golublji" genotip. Nalaz 12,9% golubova pozitivnih na prisutnost bakterija porodice Chlamydiaceae jasno pokazuje prisutnost uzročnika u jatima golubova u Hrvatskoj, te mogući rizik za uzgajivače koji dolaze u bliski kontakt s pticama.

Ključne riječi: Chlamydia psittaci, genotipovi, golub, Hrvatska